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Potassium Channel Activation: A Potential Therapeutic Approach?

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ABSTRACT. The physiological role of K* channel opening by endogenous substances (e.g., neurotransmitters and hormones) is a recognised inhibitory mechanism. Thus, the identification of novel synthetic molecules that 'directly' open K* channels has led to a new direction in the pharmacology of ion channels. The existence of many different subtypes of K* channels has been an impetus in the search for new molecules demonstrating channel and, thus, tissue selectivity. This review focuses on the different classes of openers of K* channels, the intracellular mechanisms involved in the execution of their effects, and potential therapeutic targets. Pharmacol., THER. 78(1): 39-63, 1996.

KEY WORDS. Porassium channel openers, KCO classification, KAII, KG, cromakalim, therepount targets.

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ABBREVIATIONS. BK_{Ca} high-conductance calcium-activated potassium channel; [Ca⁷⁺], Intracellular Ca⁶⁺ concentration; cAMP, cyclic AMP; cOMP, cyclic GMP; CORP, calcitorin gene-related peptide; DH5-1, dehydroso-yassponin li DMPP, 1,1-dimethyl-4-phenylpiperannium; EDHP, endothellum-derived hyperpolarizing factor, EDRF, endothellum-derived relaxing factor, E₂, potassium equilibrium potential, ET-1, endothellum-1; 5-ET, 5-hydroxytrypramine; K_{AB}, ATP-sensitive potassium channel; K_{Ca}, calcium-activated potassium channel opener; KCO, potassium channel opener; K_{CB}, ATP-sensitive potassium channel opener; CO, ATP-sensitive potassium channel opener; K_{CB}, conductance opener; K_{CB}, CD, at a potassium channel opener; NDP, nucleotide diphosphate; NO, nitric oxide; PG1, prostacyclin; pS, conductance; SK_{Cb}, small-conductance calcium-activated potassium channel; SMC, smooth muscle cells; VOC, voltage-operated Ca²⁺ channel.

1. INTRODUCTION

The biological cell is an integral structure that responds to chemical and physical extracellular signals on its membrane, which are communicated to the intracellular processes through a variety of pathways. The transmembrane movement (i.e., efflux and influx) of ions (e.g., Ca²⁺, Na+, K+, Cl⁻¹) through plasmalemma channels are universal mechanisms used to execute or modulate physiological functions in living cells.

Potassium (K+)-specific channels are a diverse and ubiqzirous group of ion channels and, thus, play a fundamental role in the modulation of cell excitability (Hills, 1984; Rudy, 1988). In the resting state of excitable or nonexcitable cells, the concentration of K+ outside the membrane (3-5 mM) is at least 25-fold lower than the K+ concentration in the intracellular fluid (130-160 mM). Consequently, an outward current due to efflux of positively charged ions is generated by the opening of K+ channels. The efflux of K+ is a mechanism for recovering (repolarization), maintaining (clamping), and/or enhancing (hyperpolarization) the resting potential of the cell. Thus, the opening of K+ channels is a physiological means for counteracting, rescricting, or preventing depolarizing activity caused by inward currents, due to entry of Ca²⁺ and Na+ and the efflux of Ci¹⁺ ions.

The functions of these charmels, which are crucial for subserving different physiological functions, depend on the

specific manner in which a particular K+ channel opens and closes, and its selective permestion by K* ions. K* channels are generally classified according to their primary regulatory or gating mechanism (Hille, 1984). Ion channels can be characterised by ionic selectivity (differential permeability), conductances (p5), gating properties (factors controlling channel opening and closing), kinetics (rates at which channels open and close), and charmacology (action of specific agents in blocking or changing the flow of ions). K* channel classifications and their phurmacological properties have been reviewed extensively (Rudy, 1988; Casile er al., 1989; Cook, 1990; Drever, 1990; Koth, 1990; Robertson and Steinberg, 1990; Aronson, 1997). This article, however, will focus on the subtypes of K' channels (e.g., ATP-sensitive potassium channel (KAD), calcium-activated potassium channel (Kral) that have been associated with synthetic and endogenous openers of those channels.

The discovery and development of selective ligands that interact with specific K* channels, together with the combination of recent electrophysiological and molecular hiology techniques, has resulted in a more detailed characterization of the role these channels play in regulating cell function. An appreciation of the primary amino acid sequence of each K* channel protein, through molecular biology techniques, will allow the development of highly selective ligands that open a designated channel subtype. Such ligands would provide essential information regarding the physiological and pathophysiological importance of particular K* channels; thus, leading to the identification of drugs focused for defined clinical conditions.

The main objective of this review is to present the different classes of agents that open K* channels and discuss potential clinical targets.

2. POTASSIUM CHANNEL OPENER CLASSIFICATION

The emergence of synthetic potassium channel openers (KCOs) has only occurred recently; however, the physiological role of Ko channel opening by endogenous substances (neurotransmitters and hormones) is a recognized inhibitory mechanism (Kurachi et al., 1986; Bulbring and Tomita, 1987). The reem "porassium channel openers" was introduced to describe a group of novel synthetic molecules, typified by cromakatim (Hamitron et al., 1986; Hamilton and Weston, 1989), that have led to a new direction in the pharmacology of ion channels. Hamilton e. al. (1986), reporting that cromukalim evoked smooth muscle relaxant effects by the opening of K+ channels in cell membranes, initiated major research efforts in the search for other such molecules and in the determination of the specific channel(s) involved. ECO properues have, subsequently, been demonstrated in a diverse range of synthetic chemical structures and endogenous subscances. In addition, the existence of so many different subtypes of K+ channel has been an impetus in the search of new molecules that would have profiles and channel selecriviries different from those exhibited by the group of KCOs typified by cromakalim. Recent progress has been reported in the search for agents that 'selectively' open calciumactivated K* channels (see Section 2.2). In addition to the development of synthetic molecules, a number of endogetions substances have been identified that exert some of all of their effects via K* channel opening (see Section 2.3).

The recent advances made in this area of research have brought into question the use of the broad turm "potassium channel openers" for reference to a limited group of molecules (i.e., Kare openers, those typified by cromakalim). For the purpose of this review, the term "potassium channel opener" (and the abbreviation KCO) will be used in reference to any substance reported to open K* channels and, thus, associated with an efflux of K* ions from the cell. A subscript to the K of the abbreviation KCO will be used to indicate the postulated channel involved (e.g., Kari-CO for openers of Kari-channels).

Molecules (primarily KATPCOs) exhibiting properties comparible with the opening of K* channels have been essential tools in the development of research approaches and pharmacological rechniques within this area. Such techniques not only have assisted in the identification of other KCOs, but are crucial in the determination of the mechanism(s) of action and K* channel subtype involved.

The unidirectional movements of specific ions can be assessed by isotope flux techniques. The efflux of K+ from a cell can be measured by using **K+ or the more stable tracer **Rh+. Briefly, the rissue or cells are loaded with the isotope during an incubation period (Quast and Baumin, 1988). The preparation is then challenged with a KCO and the radioactivity within the bathing environment and the residual radioactivity in the rissue is determined, thereby giving qualitative and quantitative representation of K+ ion movements. In experimental models where mechanical responses can be recorded, the K+ flux can be compared with functional responses following exposure of the tissue to the KCO.

KCOs hyperpolarize the cell membrane up to values approaching the potassium equilibrium potential (Ex; see Section 31). Electrophysiological techniques permit the recording of transmembrane loss movements and membrane potential, depending on the experimental methodology, at a high level of resolution. Parch-clamp procedure applied to whole-cell confirmation allows macroscopic K* cuttents to be recorded when depolarizing pulses are delivered to the preparation clamped at negative potentials close to be (Hamill et al., 1981). Recording from isolated membrane parches also provides measurements of electrical currents through single K+ channels (Hamil et al., 1981). The difficulty in identifying certain K* channel subtypes in some tissues using these exchangues may be due to their low density, fragility to the isolation procedure, and/or their irreversible randown (where the activity of the K. channels recorded in cell free patches spontaneously declines over a relatively short period) in the absence of the cytosol. Thus, experimental conditions, such as the isolation procedure and recording techniques, can influence the determination of the properties of K* channels.

Radioligand binding assays have identified binding sites

for selected KADCOs, [HIP1075 (pinacidil analog) and [44]gromakalim, in smooth muscle preparations (Howlett and Longman, 1992; Quast et al., 1993). [PH]P1075 binding in rar acita was inhibited by representatives from all chemical families of KATPCOs with potencies that correlated with the potencies obtained in MRb* efflux and vasorelaxation studies (Quast et al., 1993). Differences in the data obtained with [HIP1075 and [Hi]cromakalim in smooth muscle preparations, however, suggest the existence of two different binding sites for KATECOs (Lawson and Hicks, 1993). The use of strips of tissue with intact cells, not membrane prepararions, being critical in this assay would suggest that the binding of these KanCOs is dependent on the functional integrity of the ceil (Quast et al., 1993). The development of similar assays for other subtypes of K* channel, however, are dependent on the availability of appropriate ligands. Ligand binding studies on expressed cloned K+ channels te.g., ROMKI; Ho et al., 1993), which should provide major advances in our understanding of how KCOs interact with the channel, are awaited.

Functional isolated organ preparations have been used extensively in KCO research to determine the mechanical effects due to exposure of tissues to such compounds. Studies in parallel with those techniques described above can demonstrate whether or not the functional responses to KCOs are compatible with and, thereby, a consequence of, K* channel opening and K* flux. Finally, the effects of KCO are determined in preclinical in wwo models predictive of potential therapeutic applications (see Section 4).

2.1. ATP-Sensitive K+ Channel Openers

KATE channels, which have been studied extensively, inirially were identified in cardiac cells and pancriatic B-cells (Norna, 1983; Asheroft and Asheroft, 1990). The intracelfular concentration of ATP determines the state (open, closed) of the KATP channel (Edwards and Weston, 1993). Cook and Hales (1984) proposed that KATF channels in pancrearic & cells are spontaneously open under normal conditions and, as a tesult, a basal efflux of K+ from the cell leads to hyperpolarization of the cell membrane. An increase in the glucose levels evokes a rise in intracellular ATP levels, closing the Kase channel. The resultant membrane depolarization and subsequent Ca2* influx through voltage-operated Ca2+ channels (VOCs) stimulates insulin release. Exogenous compounds, in particular the sulphonylureas (e.g., glibenclamide), by closing Karr channels, can induce the release of insulin from pancreatic \$\beta\cells.

Subsequent studies have shown KATE channels to exist in virtually all tissues studied, including skeletal muscle, smooth muscle and neuronal cells (Ashcroft and Ashcroft, 1990). Five different types of KATE channels have been defined as a consequence of potassium selectivity and sensitivity to calcium, intracellular ATP concentration, and pharmacological modulation (Ashcroft and Ashcroft, 1990). Full pharmacological characterization of the putative subtypes of KATE channels is presently limited (Ashcroft and Ashcroft, 1990; Gopalakrishnan et al., 1993). Of these channels,

the Type I, which are blocked by micromolar concentrations of intracellular ATP, have been extensively studied in a number of cell types. This channel, irrespective of location, always demonstrates sensitivity to sulphonylureas; however, the concentrations of these compounds moulted to block the KATE channels are tissue-dependent (Edwards and Weston, 1993). In pancrearic \$-cells, modulation of insulin release occurs at nanomolar concentrations of glibenclamide, and micromolar concentrations are required in smooth muscle and cardiac preparations, suggesting at least two concentration ranges of activity. Further, the rank order of potency of KATECOs on Type I KATE channels is dependent on the cell type being studied (Edwards and Weston, 1993) and, thus, is open to further differentiation on the basis of pharmacological profile. In addition, the current nomenclature does not take into account the results of recent studies (albeit limited) using molecular biology of the cloned KATP channel (Ho et al., 1993).

Classification of compounds termed KanCOs has largely been defined as a consequence of their biological effects being sensitive to blockade by sulphonylureas. Porassium channel opening properties, as identified by electrophysiological and efflux studies, sensitive to glibe nolamide (or other sulphony)ureas) have been demonstrated in a diverse range of synthetic chemical structures (benzochiadiazines [e.g., diazoxide], pyrimidines [e.g., minoxidil], pyridylcyanoguanidines [e.g., pinacidill, nicorinamides [e.g., nicoraudil), benzopyrans [e.g., cromakalim] and carbothiamides [e.g., RP 49356]) (Edwards and Weston, 1990; Lawson and Hicks, 1993). Common chemical atructural features between behappyrans, pyridylcyanoguanidines, and carbothiamides have been described (Atwal, 1992). In addition, most of the biological activity resides predominantly in the (--)-enantiomers of cromakalim, pinacidil, and RP 49356.

In functional in vitro pharmacology studies, the antagonism by glibenclamade of the effects of KaniCOs on vascular (Eirre, 1989a; Wilson, 1989; Newgreen et al., 1990) and certain nonvascular (Elize, 1989b; Piper et al., 1990; Edwards et al., 1991) smooth muscle preparations has been described as competitive in nature. Competitive aniagonism is inferred by parallel displacements of the KateCO-induced relaxate concentration-response curves to the right of comrols, without a reduction of the maximum effect to the 'agonist' by the antagonist, and Schild analysis (Arunlakshana and Schild, 1959) yielding a slope not different from unity. The failure of gliberulamide (up to 10 aM) to displace [H]cromakalim from its binding sites on 1st acres (Howlest and Longman, 1997) does not support a competitive interaction between these two compounds at a single site. Olibenclamide concentration dependently increased the dissociation rate of the [HIP1075 binding complex on rat sorts (Quest et al., 1993), suggesting that the glibenclamide site is distinct from, but negative allosterically coupled to, the binding site for the openers.

Parallel displacement of concentration-response curves could also be observed with physiological antagonism and, also, if spare channels were available (i.e., the KATPCO can

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evoke a maximal response without the activation of all of the channels in the tissue). The smooth muscle relaxant responses to KatpCOs in the guinea-pig traches, however, are blocked by gliberidamide in a manner that is not consistent with competitive antagonism (i.e., the maximum effect of the KatpCO is reduced in the presence of the antagonist) (Nielsen-Kudsk et al., 1990; Berry et al., 1991; Small et al., 1992), but is consistent with a lack of spare channels (i.e., the KatpCO must activate all channels in the rissue to evoke a maximal response). Thus, glibenciamide appears to interact differently with the channel activated by KatpCOs in respiratory smooth muscle from that in vascular rissue.

In vascular preparations, glibenclamide anragonized the effects of cromakalim competitively, whereas the effects of minoxidil sulphate were attragonized in a noncomperitive manner (Wickenden et al., 1991). These findings led to the suggestion of the existence of different subtypes of K* channel sensitive to KarcCOs. In addition, minoxidil sulphate, in contrast to diazoxide and cromakalim, had no effect on "Rb* efflux in rat sorts, but did increase *K* efflux (Newgreen et al., 1990; Wickenden et al., 1991). All three KatcCOs increased "Rb* and *K* efflux from rat portal vein (Newgreen et al., 1990). Thus, in rat sorts, minoxidil sulphate may open a K* channel impermeable to *Rb* and sort recognised by other KatcCOs.

A number of anomalies exist within the pharmacology of the different chemical structures that have been remoously classified into the family of KAR openers (Lawson and Hicks, 1993). A common leagure responsible for the grouping of these compounds has been the susceptibility of the hiological effects to inhibition by salphonylureas. Nevertheless, the tank order of potency of the inhibitory activity and nature of annagonism by the sulphonylures is dependent on the cell type and KarrCO being studied (Longman and Hamilton, 1992; Edwards and Weston, 1993; Lawson and Hicks, 1993). The pharmacological profile of (certain) KapCOs on the Kair channel is also dependent upon cell rype (e.g., diazoxide is an 'activator' in pancreatic cells, inactive in skeleral muscle cells, and an lantagonise in cardiac cells) (Zunkler et al., 1988; Faivre and Emdlay, 1989; Weik and Neumcke, 1990). Discrimination between the pharmacology of benzopyran and nonbenzopyran KATCOs has been identified (Lawson et al., 1992; Randall and Oriffith, 1993), where all compounds have exhibited definable KCO profiles. Such findings support the concept that there are differences in the way these agents interact with pointsium channels, whichever subtype they may be (Lawson et al., 1992; Lawson and Hicks, 1993).

The ability of glibenclamide to entagonize the actions of this group of compounds led to the suggestion of Kate channels being the site of action; however, this is still an area of research and discussion. Determination of a common site of action for this group of compounds is complicated by, at least, (I) the existence of subtypes of KATP channels, (2) evidence of these compounds opening several K* channels, and (I) effects of gliberclamide on K* channels other than Kate channels.

Using parch-clamp techniques on the rabbit portal year, Beech et al. (1993a,b) found that nucleotide diphosphates (NDPs), such as CDP, are required for Kate channel opening in the absence of the inhibitory effects of ATP. This propcity sets them aport from the Kary channel present in the pancreas and heart. These authors suggested that this K* channel in rabbit portal vein should be more correctly termed Know to reflect the obligatory role that NDPs play in regularing this particular channel. Together with the recent hadings of Kitomura and Kamouchi (1993) that Kare channels appear to be nonuniform across species in various smooth muscle cells (SMC), evidence suggests that there may be several different types of ATP-, adening nucleotide, and gliberalismide-sensitive KATP channels in various tissues that may be important targets for compounds that can selectively regulate their activity. These findings highlight the complexity of the biological system and the need to fully investigate the specificity of channel/ligarid interactions to clearly define the size of action of openers of Karr channels.

A number of KARCOs, however, (e.g., pinnall and nicorandil) possess additional properties (e.g., adonylate or guanylate cyclase stimulation) to that of porassium channel opening that could account, in part, for pharmacological profiles differing from those of the bencopyrans (e.g., cromakalim) and carbothiamides (e.g., RP 49356) (Cook, 1990; Longwan and Hamilton, 1992). The situation is further compheated by agents defined as KaryCOs and glibenclamide exhibiting the actions on K. channels other than KAD channels, Care channels and Ct. currents. Glibenclamide can inhibit delayed rectifier K+ currents (Reeve et al., 1992; Berch et al., 1993h, Grepel et al., 1993), A type K*-currents (Sadraei and Berch, 1993), Cal+ channel currents (Bion and Hormsmeyer, 1994; Sadraei and Beech, 1995) and Cyvic Fibrasis Transmembrane Regulator-dependent CI - current (Sheppan) and Welsh, 1992). Cromakalim activated Kis, current (Oelband and McCullough, 1993) and inhibited Culchannel current (Okabe et al., 1990) and Cystic Fibrosis Transmembrane Regulator-dependent Cl. current (Sheppard and Welsh, 1992).

A hererogeneity in the pharmacology of the KateCOs can be identified, which would suggest a heterogeneity of the Kory sites of action and/or the K* channel. Interpretadon of dara is further complicated by the potential beterogenerry of sulphonylurea-sensitive sucs of deactivation that may be independent of the KALECO site of action. Further work involving comparative studies with representatives of each chemical family of KatiCOs will assist in the understanding and subclassification of these hererogeneous sites of action. A combined effort of electrophysiology, huncrional pharmacology, and molecular biology, with a subsequent study of the expressed clone(s), will be required to address the real question of channel selectivity and Kan CO subclassification. The differences already observed to the pharmacology of KANCOS me, therefore, important factors to consider in the development of second generation compounds, where rissue or organ schemivity is sought

2.2. Calcium-Activated Potassium Channel Openers

Ken channels are activated by membrane depolarization and by increases in intracellular calcium (Blatz and Maglety, 1987). Three subtypes of Ko, channels have been described on the basis of their single-channel conductance and senshivity to specific pharmacological blockers (Cook, 1990). High-conductance or maxi-K (BKCai, 100-250 pS) channels are sensitive to charyhdotoxin and iberiotoxin, intermediateconductance (IKCs; 18-50 pS) channels are blocked by high concentrations of charybdotoxin, and low or smallconductance (SK_{Cs}; 10-14 pS) channels are potently blocked by apamin Ko, channels have been identified in virtually all types of cells, where they almost certainly function to terminate excitatory processes that are triggered or maintained by an increase in the intracellular Ca2+ concencration ((Cal*)) and/or involve depolarization. As a consequence of such an important physiological tole of KCs channels, the search and recent identification of selective symberic BKCs openers has received significant attention.

Two major chemical groups, benzimidazoles (e.g., NS-004, NS-1619) and imidato(1,Z-a)pyratines (e.g., SCA40), recently have been reported to exhibit BKC, channel opening properties (Sablayrolles et al., 1988; Oleson and Watjen, 1992; Laurent et al., 1993; Olesen et al., 1993). NS-004 directly acrivated the charybdorexin-sensitive BKCa channel in rat cerebellar granule cells, in rat GHI clonal pituitary tumor cells, bovine aomic SMC and guines-pig tracheal SMC (Olesen and Watjen, 1992; Olesen et al., 1993, 1994a; McKay et al., 1994). NS-1619 directly activated the BKCs channel in rat ventremedial hypothalamic neurones, bovine actic SMC, and rar portal vein (Edwards et al., 1994; Sellers and Ashford, 1994; Olesen et al., 1994b). Although NS-1619 failed to modify the Extendence in rat ventromedial hypothalamus (Sellers and Ashford, 1994), in the rat portal vein, this compound was reported to inhibit the KV channel over the same concentration range that activated the BKC, channel (Edwards et al., 1994). NS-004 induced relaxation in guinea-pig trachea, which were sensitive to charybdotoxin and therlotoxin, but not glibenclamide (Karr channel), dolenlide (inward rectifier) or spamin (SKOs), suggesting activation of only B&3 channels being responsible for this functional response (Lawson et al., 1996). Although NS-004 and NS-1619 activate BKC, channels, their relaxant effects in rat portal vein and cardioprofective effects probably are due to inhibition of L-type calcium channels (Edwards et al., 1994; Sargere et al., 1993).

In guinea-pig tracheal amoorh muscle, SCA40 evoked relaxations that were inhibited by charybdotoxin, suggesting the involvement of BKCs channels (Lautent et al., 1993). However, SCA40 had no effect on the activity of BKCs channels in bovine tracheal SMC (Macmillan et al., 1995). Cook et al. (1995) reported that relaxant effects of SCA40 in guinea-pig tracheal smooth muscle are due to the inhibition of phosphodiesterase. Activation of KCs channels in respiratory smooth muscle following \$\beta\$-adrenoceptor stimulation appears to be caused by phosphorylation mediated by a cyclic AMP (cAMP)-dependent protein kinase (Kume

et al., 1989). Thus, the K_C, potassium channel opener (K_CCO) profile of SCA 40 may be due to an indirect action on the cAMP pathway.

Dehydroso-yasaponin-I (DHS-I), derived from the medicinal herb Desmodium adsorders used therapeutically in the treatment of dysmenorrhea and asthma (Ampofo, 1977), opens charybdotoxin-sensitive K_{Cx} channels in bovine tracheal smooth muscle membranes (McManus et al., 1993). Data regarding the pharmacology of DHS-I is limited and, therefore, it is not known if it is the active factor of the herb and, if so, whether K* channels are the primary site of action.

The thiazide diuretic hydrochlorothiazide evokes relaxation in vascular smooth muscle due to the activation of charybdoroxin-sensitive K* channels (Calder et al., 1994). These effects have been associated with an increased *Rb* efflux (Calder et al., 1994) and a decrease in [Cal*], (Picklers and Hughes, 1995). An increased *Rb* efflux has also been observed with the thiazide drug cyclothiazide in arterial smooth muscle (Moura and Worcel, 1983). Whether or not thiazides directly act on a K* channel remains to be established.

2.3. Endogenous Openers

Endogenous openers of porassium channels can be divided into three basic groups: (1) ligands that modulate the gating of potessium channels following interaction with plasmalemma receptors, (2) imracellular second messengers (e.g., inositol trisphosphate, inusited 1,3,4,5 tetrakisphosphate, cAMP), and (3) extracellular substances believed to act directly on the channel. Although it is this latter group that will be the subject of this section, the former groups are of great interest in the understanding of the pathophysiology of disease states and subsequent treatment. In pathologic situations, there may be a defect in the neurotransmitter or receptor (e.g., noradrenaline, 5-bydroxytrypsamine [5-HT], acetylcholine, opioid peptides), resulting in decreased activation of the associated K+ channel. Direct modulation of the K* channel through specific agents could provide focused thempy, depending on the distribution of that channel subtype.

Endothelial cells, which line all blood vessels and the cardiac cavity, have a central role in cardiovascular homeostasis, at least in part, through the release of substances that mediate control of vascular tone and cardiac contraction (Shah, 1992). Stimuli of receptors (e.g., acetylcholine, bradykinin) or nonreceptot processes (e.g., electrical field stimulation, calcium ionophote A23187) induce endothelium-dependent hyperpolarization of SMC, leading to vasorelaxation (Garland et al., 1995).

The first endothelium-derived mediator found to act on vascular SMC was prostacyclin (PCI₂), a vasorelaxing cyclo-oxygenase product (Weksler et al., 1978). The stable analog iloprost, like prostacyclin, evokes a hyperpolarization of SMC through the activation of K* channels (Siegel et al., 1990). K_{Cs} channels have been proposed to be the site of action

of iloprost (Boeynaems and Ramboer, 1990); whether this property of iloprost is due to activation of POI₂ receptors or a direct action on the K⁺ channel is still a subject of debate. The development of other stable analogs of POI₂ that demonstrate therapeutic benefit (e.g., in peripheral arretial occlusive disease, Raynaud's syndrome, cardioprotection, stroke) by the activation of K⁺ channels is an active area of research.

Furthgort and Zawadski (1980) demonstrated the imporcance of the vascular endothelium in mediating the relaxant effect of acetylcholine, leading to the proposal of the existence of endorfuelium derived relaxing factor (EDRF), EDRF is believed to be nitric oxide (NO) or a closely related NOreleasing molecule (Palmer et al., 1987). EDRF-induced smooth muscle relaxation is mediated, at least in pact, by the haem-dependent acrivation of guanylate cyclase, with the subsequent generation of cyclic GMP (cGMP) (Furthgott and Jothianandan, 1983; Rapoport and Murad, 1983). Before the discovery of the role of the endorhelium in vasodilacion, Kuriyama and Suruki (1978) had observed that acetylcholine hyperpolarized SMC by increasing the membrane permeability to K* ions. Although NO has been reported to change smooth muscle membrane potential (Tacc et al., 1990) and directly accivate KCs in SMC (Bolotina et al., 1994), the contribution of these effects to endothelial dependent relaxation relative to direct stimulation of guarrylare cyclase is unclear. Taylor and Weston (1988) subsequently suggested that an additional factor to EDRF, which could cause vasorelaxation by increasing the membrane potential of the muscle cells, was released from the endothelium by acetylcholing. To distinguish this factor from EDRE, it was remed endothehum-derived hyperpolarizing factor (EDHF). Although evidence has been presented to support the hypothesis that EDHF is a diffusible factor released from the endothelium, an electroussic spread of hyperpolarization between the endothelist and SMC cannor be eliminated ar this stage (Garland et al., 1995). The chemical identity of EDHF remains unresolved; however, a number of candidates (e.g., prostanoids) have been eliminated (Carland et ul., 1995). The proposal that EDHF may be a cytochrome P450-derived arachidonic acid metabolite, such as epoxyelcosarrienoic acids, requires confirmation (Gebremedhin et al., 1997; Hecker et al., 1994). Experiments to advance the identification of EDHF may be complicated by the existence of a family of factors, as opposed to a single substance.

The effects of EDHF are associated with the efflux of K* ions from SMC (Chen and Suzuki, 1989), but the channel(s) involved have not been definitively characterized. The majority of studies have found that glibendamide blocks the change in membrane potential, suggesting the involvement of K_{ATF} channels (Chen and Cheung, 1992; Eckman et al., 1992, Garland and McPherson, 1992; Adeagho and Triggle, 1993). In certain vascular beds, however, K_{Cr}, channels have also been proposed to be the site of action of EDHF (Adeagho and Triggle, 1993). The existence of more than one EDHF may also complicate the identification of the site of actions.

Physiologically, the action of a hyperpolarizing factor that

is distinct from NO appears to predominate in the mediation of endothelial-dependent smooth muscle relaxation in small resistance arteries (Garland et al., 1995). In large afteries, both NO and EDHF appear to contribute to relaxation, with NO being dominant under normal circumstances. Thus, EDHF and the K+ channel it activates may be of primary importance in the regulation of vascular resistance. The clinical importance of EDHF in disease actiology will only be appreciated when the chemical identity is established and/or the K+ channel(s) activated is characterized.

The endothelins are a family of 21 amino acid vasoacrive pentides, of which endothelin-I (ETI) has been reported as the most porent vasoconstrictor known (Randall, 1991). ET1, however, has also been demonstrated to modulate Kapp channels in both in view cell culture (Inone et al., 1990) and in vivo studies (Hasunuma et al., 1990; Lippton et al., 1991). As a consequence of this property, ETI preferentially evokes vasodilation in certain vascular beds (e.g., pulmonary; Lippron et al., 1991). In addition, the vascular effects of benropyran, but nor carbothismide and oyridyle yanoguanidine, KangCOs (see Section 2.1) were modified by ET-1, suggesting that BRL-38227 and ET-1 have affinity for a common site on the Karr channel (Lawson et al., 1992). This bypothesis is supported by the hading that BRL-38227 inhibits binding of [13 IEFI to ret cardiac membranes (Waught et al., 1992). Further, ETI evokes membrane hyperpolarization in rat antic SMC (Van Renterghein et al., 1988), for glioma C6-BU-I cells (Cleason et al., 1991), porcine cotonary arresty cells (Hu et al., 1991), rat gastrat longitudinal and circular smooth muscle (Fulginia et al., 1993) and goineapig menia coli (Usune et al., 1991) through the activation of charybdotoxin-sensitive KC, channels. In contrast, ET-1 also exhibits antagonistic properties at both Kate (Miyoshi et al., 1992) and BKC, (Hu et al., 1991) channels

Consequency, EFI can directly modulate K* channels having affinity for both K_{KD} and K_{Cs} channels. The (patho)physiological role of EFI-induced K* channel activation remains to be established. The recent advances in the identification of selective antagonism will allow the elucidation of the involvement, if any, of ET-receptors in this action of the peptide.

Interestingly, ET4 and ET3 can also evoke an indirect endothelium-dependent hyperpolarization in car mesenteric smooth muscle (Nakashima and Vanhourte, 1993a), but not in canine coronary arrery (Nakashima and Vanhouste, 1993b).

Finally, a variety of endogenous polypeptides have been proposed to exhibit K+ channel opening properties. The endogenous vascalilator peptides, vascactive intestinal peptide and calcironin gene-related peptide (CGRP), activate KALP channels in vascular SMC, leading to hyperpolarization and tissue relaxation (Standen et al., 1989; Nelson et al., 1990), CGRP is a polypeptide located in neurones that form a close association with both central and peripheral blood vessels (Bevan and Brayden, 1986). The CGRP-induced relaxations of some blood vessels (e.g., rat sorta) are endothelium-dependent; thus, the role of endothelium-dependent; thus, the role of endothelium-

derived factors in the observed hyperpolarization has been proposed (Grace et al., 1987; Nelson et al., 1990). Galanin and somatostatin, hyperglycemic hormones, activate K_{ATP} channels in insulin secreting pancreatic β-cells (De Weille et al., 1988, 1989). The block of anoxia-induced depolarization of hippocampal CA3 neurones and glutamate release inhibition by galanin is consistent with K⁺ channel opening (Ben-Ari, 1990). However, not all reports support the conclusion of K⁺ channel activation as a property of vaso-active intestinal peptide, CGRP, galanin, and somatostatin; further work is required to establish the role of K⁺ channels in their physiological profiles.

2.4. Others

1,1-Dimethyl-4-phenylpiperazinium (DMPP), a selective nicotinic receptor stimulant, has shown activity consistent with K* channel activation in the isolated tunica muscularis mucosae of the rat oesophagus (Adeagbo et al., 1993). Neither apamin nor glyburide modified the DMPP-induced relaxation. Whether this was a direct action of DMPP on a K* channel or a nicotinic-cholinoceptor-linked K* channel requires elucidation.

3. MECHANISM(S) OF ACTION

The standard criteria that initially identified a compound as a KCOs has been its ability to relax an in vitro smooth muscle preparation contracted with low, but not high, concentrations of extracellular K* Jone (Weir and Weston, 1986); Bray et al., 1987; Hamilton and Weston, 1989; Lawson and Cavero, 1989). The principal determinant for inhibition or reversal of smooth muscle contraction by a KCO is a reduction in the free cytosolic Ca3* concentration. The mechatilsm(s) involved in the production of smooth muscle relaxarion by K+ channel openers, especially KanCOs, has been (and still is) a major subject of debate. To date, information regarding mechanism(s) of action primarily has been obtained from studying KarrCOs, especially cromakalim and other behropyrans. Investigations with other classes of KCOs (e.g., KoCOs) that will demonstrate whether or not mechanism(s) of action are common between the different K* channels and how they are acrivated by different ligands, are awaited. The mechanism of action involved in a given response may not only depend on the subtype of K+ channel, but also how the KCO interacts with the channel to evoke that effect (Lawson et al., 1992; Randall and Oriffich, 1993).

Although some mechanism(s) are universal in different excitable and nonexcitable cells, not all processes will be applicable to all types of tissue. The majority of data has been obtained from SMC and requires confirmation in other cells.

3.1. Hyperpolarization

The opening of K^* channels by these compounds and subsequent efflux of K^* ions from the cytosol leads to membrane repolarization and/or hyperpolarization (Cook, 1990; Edwards and Weston, 1990; Robertson and Steinberg, 1990; Longman and Hamilton, 1992). This change in membrane potential is followed by a reduction in cytosolic free Ca²⁺ and/or an inhibition of mechanisms producing increases in cytosolic free Ca²⁺. The outcome of these effects is a reduction in membrane and cell excitability, resulting in a greater cellular resistance to activation by excitatory stimuli.

In amounth muscle preparations, the relaxant actions of $K_{APP}COs$ have been accompanied by an increase in negativity of the mating membrane potential (hyperpolarization) towards the calculated E_{K_1} together with an outward current of K^* ions (Hamilton and Weston, 1989).

An excellent correlation exists between the potencies of the KAPCOs for stimulation of *Rb* efflux and vasorelaxation in the acrta and potent vein in vine preparations (Quasterul., 1992), supporting the hypothesis that the functional responses rely on the opening of plasmalemma K* channels in vascular smooth muscle.

Although, initially, it was assumed that the hyperpolarization caused by K+ efflux produced closure of VOCs, preventing depolarization-induced Ca²⁺ entry into the cell, other mechanisms, in light of recent evidence, may also contribute to the effects produced by K+ channel openers (see Sections 3.2, 3.3 and 3.4). Evidence that cromakalim-evoked increase in ⁸⁸Rb+ efflux or hyperpolarization of vascular smooth muscle tissue was not influenced by latthanum or the Ca²⁺ antagonist nifedipine, is indicative that the action of the Kan COs is not dependent upon modifying the influx of external Ca²⁺ ions (Coldwell and Howlett, 1988; Southerson et al., 1988).

Vasorelaxant effects of Kath COs have been reported that are independent of membrane hyperpolarization or ion efflux (Hamilton et al., 1986; Quasi and Baumlin, 1988; Greenwood and Weston, 1993). Such findings suggest that rhese drugs can exert a response through mechanisms other than the opening of K* channels. The possibility that Kath COs do not interact directly with an ion channel bur, rather, for example, with an enzyme system involved in intracellular phosphotylation, provides a novel explanation for some of the apparently anomalous effects of these agents (Edwards and Weston, 1994).

3.2. Intracellular Ca2+ Stores

Experimental evidence suggests that K_{ATI}COs may also have a direct effect on intracellular stores. Cromakalim evoked a contractile response in rabbit aorta bathed in a Cai⁺-free solution, which may be related to effects on intracellular Cai⁺ stores (Duty and Weston, 1992). Using rabbit cultured tracheal SMC, Chopra et al. (1992) demonstrated that cromakalim reduced the uptake into and inhibited the release of *Cai⁺ from the sarcoplasmic reticulum. These findings support those obtained from vascular smooth muscle, where contractile responses to notathenaline, dependent on intracellular calcium stores, were ortenuated by cromakalim (Bray et al., 1991).

In contrast, the effect of cromakalim on rat pulmonary artery did not appear to involve an action on Ca^{2*}-release from internal stores (Savineau and Marthan, 1993).

3.3. Inositol Phosphare Cascade

BRL 38227 inhibited Ca²⁺ release from intracellular stores of rabbit isolared mesenteric artery due to an action on noradrenaline-stimulated 1,4,5-inositol trisphosphate production (Ito et al., 1991). This effect was sensitive to glibenciamide and high (128 mM) extracellular KCl. Hyperpolarization of the plasma membrane of canine coronary artery by KarrCOs has also been associated with an inhibition of the production of 1,4,5-inositol trisphosphate and, hence, Ca³⁺ release from intracellular stores (Yamagishi et al., 1992). Interestingly, KatrCOs have shown to effect upon phosphatidyl mositol turnover in brain tissue (Coldwell and Howlett, 1988).

3.4. Ca2+ Sensitivity

The hyperpolarization by KanCOs (e.g., levrocromakalim) may also be linked with a reduction in the sensitivity of the contractile elements of vascular smooth muscle to Ca^{7*} (Okada et al., 1993). Cromakalim, however, has no direct effect on the contractile proteins of skinned trachealis muscle (Allen et al., 1986).

3.3. Others

Cromakalim has been reported to stimulate the Na-K pump in human and canine mesenteric arreries through an elevation in intracellular Na* (Hong et al., 1993). In rat sorts, cromakalim appears to be absent of effects on the Na-K pump (Cavero et al., 1987).

The KAPCOs, cromakalim, and RP-49356 failed to increase cAMP or cGMP in SMC (Southerton et al., 1988; Nakajima et al., 1989), or to potentiate the effects of forskolin its airway smooth muscle preparations (Berry et al., 1991; Murray et al., 1990). Pinacidil and cromakalim do not appear to be inhibitors of phosphodiesterase activity in airway smooth muscle (Berry et al., 1991; Ho et al., 1990). Quanine nucleocide-binding proteins (G-proteins) are known to provide a hisk between membrane receptors and intracellular events, and the possibility exists that G-proteins may modulate the activity of a K1 channel sensitive to KAPCOs. Results with persussis or choleta toxin, however, suggest that the corresponding G-proteins are not involved in the actions of cromakalim in vascular smooth muscle (Longman and Hamilton, 1992).

Therefore, the involvement of intracellular Ca2* stores in the responses to KanCOs may be species, and/or tissue-dependent. Further studies are required to determine whether or not this is a property unique to cromakalim (or bensupyrans) and if it is a direct effect or a consequence of hyperpolarization. Thus, the mode of action of K* channel openers may not be as simple as first thought and more research effort is required in this area.

4. THERAPEUTIC TARGETS AND POTENTIAL

Theoretically, at least, the general decrease in the excirability of cells that follows K* channel opening infers a broad clinical potential for drugs with this property in a number of pathologic conditions.

4.1. Cardiovascular System

The preclinical profile of K_{AD}COs clearly supports a clinical potential for their use in vascular pathologies that require a decrease in peripheral vascular resistance, an inhibition of excessive vasoconstriction, and/or a prolongation of myocardial tissue viability, while undergoing transient oxygen deficiency.

4.1.1. Vascular. The pharmacological profiles of KATECOs. in vascular smooth muscle tissues and in view models of vascular disorders (e.g., hypertension, peripheral vascular disease, angina) have been reviewed extensively (Edwards and Weston, 1990; Richer et al., 1990; Longman and Hamilton, 1992; Edwards and Weston, 1995). In a variety of vascular tissues from a range of species, KATPOOs display the ability to relax smooth muscle by inhibiting both spontaneous tone and/or spasmogen-induced contraction. Afterial smooth muscle tone, of which K* channels in the SMC are imporsant regulators, is the main determinant of peripheral vascular resistance and blood pressure. Functional defects of K* channels may lead to vasoconstriction or compromise the ability of an arrery to dilate in pathologic conditions of the vascularure, such as vasospasm, hypertension, ischemia, hypotension folkowing sepsia, and diabetes.

Activation of K_{AIT} channels, which respond to the metabolic state of the cell, may be involved in arternal dilation in reactive hyperemia, septic shock, ischemia, and hypoxia (Nelson, 1993). The involvement of K_{AIT} channels in these clinical conditions may not be exclusive; however, the absence of selective agents of other K* channels presently limits full characterization of the actiology of such discases.

KanCOs can lower both normal and experimentally elevated blood pressure (Edwards and Weston, 1990; Richer real, 1990; Longman and Hamilton, 1992). The mechanism of this effect is an actively mediated decrease in rotal peripheral vascular resistance. Therapeutic experience with pinacidil, diaroxide, and minoxidil indicates that they also reduce elevated blood pressure in patients (Gross, 1977; Oates et al., 1977; Abulelt-Roune, 1988). Hypertensive patients created with pinaculil (10-25 mg bid) showed a marked fall in total peripheral resistance, the short duration of which was controlled through the development of sustained release formulations (Carlsen et al., 1983, 1985). The antihypertensive effects of pinacidil are accompanied by an initial reflex tuchycardia and by weight gain in approximately 10% of parients. Although the antihypertensive effects of diazoxide have been known for many years (Gross, 1977; Oates et al., 1977), increased blood glucuse levels has restricted its use to hyperrensive emergencies. The extent to which the antihypertensive effects of diagoside result from K+ channel opening action is still unknown, as evidence of additional effects have been observed (Newgreen et al., 1990). The relatively high incidence of fluid retention has severely restricted the clinical use of minoxidil (Gross, 1977; Oates et al., 1977). The K** channel opening profile of minoxidil, however, is different from that of diazoxide and cromskalim, whereby the involvement of a minoxidil-selective channel has been proposed (Newgreen et al., 1990; Lawson and Hicks, 1993).

Olinical experience with cromakalim is less extensive than for the above $K_{AD}COs$, and there are no published reports of clinical trials with the other KAIRCOs (e.g., RP 49356, Ro 31-6930, EMD 52692, KRN 2391). In mild to moderate hypertension, cromakalim (0.5-1.5 mg p.o.) lowered systolic and dissrolic blood pressure following a single oral dose (Vanden Burg et al., 1986; Singer et al., 1989) or chronic once-daily administration (Eckl and Greb, 1987; Lebel et al., 1989; Vanden Burg et al., 1987), with no such effects in a parallel normogensive group or when placebo was administered. As a direct vasodilator, the use of KCOs as monotherapy to reduce blood pressure can produce a series of undesirable effects (e.g., cachycardia, headache, flushing, increase in renin, aldosterone, and catecholamine secretion, and sodium and water retention (Cluck et al., 1987; Lijnen et al., 1989) that are not acceptable in clinical practice. Kan COs, however, could become useful antihypertensive therapy if appropriately formulated and coprescribed with selected agents to reduce or prevent the undestrable events. Due to apparent cardioprotective properties (see Section 4.1.2), small doses of KARCOs could be used to provide invocardial protection to hypertensive patients, an area where current drugs do not appear to substantially reduce cardiovascular mortality and morbidity (Escande and Cavero, 1992; Cavero and Premmereut, 1994; Grover, 1994a.h).

In preclinical studies, KapCOs relaxed coronary conducrance arreries, increased selectively coronary blood flow and antagonized the vasoconstructor activity of a large number of excitatory stimuli (Longman and Hamilton, 1992). If similar findings are reproduced in human subjects, Kati COs would have antianginal activity at doses that do not provoke undesirable, reflex-mediated activation of the sympathetic system, which is a deferetious physiological reaction for myocardium frankly ischemic or lacking a safety margin of blood flow reserve. Thus, these agents demonstrate propgrass desirable to improve oxygen delivery and also reduce caygen consumption within ischemic regions of patients with transient and chronic heart disease (e.g., angina pectoris). Reports on the effects of ECOs in angina pectoris, however, are restricted primarily to trials in which nicorandil was used (Flohmann, 1983; Kinoshira and Sakai, 1990). The clinical benefits of nicorandil probably result both from K+ channel opening properties and its ability to stimulare smooth muscle guswyłate cyclase (Hamilton and Weston, 1989). Cromakalim may also be beneficial in the treatment of angina pectoris (Thomas et al., 1990).

One therapeutic approach to congestive heatt failure, although it is of a symptomatic nature, is to reduce peripheral vascular resistance, a property demonstrated by

K_{ATP}CO₅ (Gopalaktishnan and Triggle, 1990). In an *in viso* rat cardiac failure model, down-regulation of ventricular K_{ATP} channel density, together with 1,4-dihydropyridinesensitive Ca⁴⁺ channels and β-adrenoceptor densities, was observed, and these changes have been implicated in this pathophysiological condition (Gopalakrishnan et al., 1990). No clinical data for the effectiveness of KCOs in the treatment of congestive heart failure has been published.

Activation of K* channels can improve the energy metabolism and the mechanical performance of skeletal muscles suffering oxygen deholency (Cook and Chapman, 1993). This is achieved partly by a selective dilation of collateral vessels supplying the ischemic skeletal muscle and, partly, by a better utilization of high energy phosphates. In rat skeletal muscle, Angersbach and Nicholson (1968) demonstrated that KADOOs, but not Call antagonists or hydralazine, selectively increased blood flow to collareral vessels in a previously ischemic limb, despite a reduction in basal diastolic blood pressure. These mechanisms are evidently of cherapeutic perential for treating patients with peripheral vascular disease, a disabling old-age disorder characterized by poor blood supply to the limbs due mosily to arberosclerosis. The role of K* channel activation in the beneficial effects of floprost (see Section 2.3; Pessi et al., 1986; Muller Buhl et al., 1987; Oberender et al., 1989) in peripheral arterial occlusive disease still remains to be determined.

Cromakalim (10 µM) increased ³⁶Rb* efflux in control cultured arterial SMC, but was without effect in cholesterol-enriched SMC (fulenko and Bialecki, 1989). Cholesterol enrichment of SMC membranes can severely influence the cellular responses to cromakalim. Thus, the benefit of KanCOs in clinical conditions associated with excess lipids may be questionable.

Kc, channels appear to play a fundamental role in regulating the degree of intrinsic tone in resistance arteries (Nelson, 1993) and, as such, help regulate arterial responses to pressure and vasoconstrictors. Therefore, activation of these channels should contribute to vasodilation. Defects in Kck channels could lead to, or contribute to, pathologic conditions that are characterized by highly constricted arteries. Thus, the development of activators may be useful in the treatment of, for example, coronary or cerebral vasospasm. Although NS-004 and NS-1619 activate BKc, channels (see Section 2.2), their relaxant effects in (at postal vein (Edwards et al., 1994) and cardioprotective effects (Sorgent et al., 1993) are probably due to inhibition of L-type calcium channels.

As blood flow increases through a conduit artery, the vessel dilates (Hilton, 1959) by an endorhelium-dependent mechanism (Hull et al., 1986; Pohl et al., 1986). Plow in rabbit isolated iliac arteries appears to activate a charyhdomnin-sensitive K* channel on the endorhelial cell membrane that leads to the release of NO (Cooke et al., 1991). Neither glibenclamide (KATE channel blocket) not apamin (SKCs channel blocket) had any effect on the flow-mediated vasodilation. Thus, in the regulation of arterial rone, BKCs channels act as the transducer of the flow stimulus, whereas NO is the effector of the vasodilation.

Endothelium-dependent vasodilations have also been associated with the activation of KATP channels. The cromakalim- and pinacidil-induced dilation of canine large coronary arteries in viso, an indirect flow-mediated effect, are entirely dependent on the endothelium (Drieu La Roschelle et al., 1992; Ghaleh et al., 1995). The KATPCO, levroctomokalim, was more potent as a vasorelaxant in rat aortic ring preparation with endothelium than in denuded tissues, on effect that involved NO (Lawson et al., 1993b).

In porcine acric endorhelial cells, pinacidil and cromakalim elevated [Ca²⁺], by inducing membrane hyperpolarization following K* channel activation (Luckhoff and Busse, 1990), an effect that can promote Ca²⁺ dependent formation of EURE Pinacidil also opened K_{ATP} channels in both rat acrta and brain microvascular endorhelial cells (Janigro et al., 1993). These findings suggest that K_{ATP} channels may play a role in the regulation of endothelial cell resting membrane potential, for example, during impaired energy supply and, therefore, modulate release of endothelium-derived vascactive factors and blood flow.

4.1.2. Cardiac. K* channel opening properties are desirable for therapeutic agents aimed at treating patients with transient and chronic heart diseases. K***ar**COs have demonstrated cardioprotective effects as a consequence of improving oxygen delivery and reducing oxygen consumption within the ischemic region (Escande and Cavero, 1992; Grover, 1994a;b; Yao and Gross, 1994; Orover et al., 1995). Both antiarrhythmic and proarrhythmic properties have been reported in K***ar**COs, leading to the safety of these drugs being a major subject of discussion (Carlsson et al., 1991; Tosaki et al., 1993; Black and Lucchesi, 1994; D'Alonzo and Grover, 1994; Wilde, 1994). Alrhough there appear to be good theoretical arguments as to why K***ar**pCOs may be of use in the treatment of some arrhythmias and in ischemic heart disease, there are major hurdles to overcome.

Clinical evidence to establish the benefits of K_{ATC}COs as treatments of patients with heart disease is awaited. Therefore, only the basic concepts of K_{ATC}CO-induced beneficial or undestrable cardiac offects will be outlined.

Cardiac KATE channels have been shown to open in response to ischemia (Kantor et al., 1990). The depletion of ATP in the myocardism and subsequent opening of Karr. channels may lead to a rapid reduction in contractility of the ischemic myocardium to protect against further ischemic injury. In support of this hypothesis, cromakalim, RP 52891 (aprikalim), and pinacidil, in animal models, cause a glibenclarifide sensitive reduction in this severity of ischemic/ reperfusion injury of the myocardium (Richer et al., 1990; Auchampach et al., 1992; Escande and Cavero, 1992; Grover, 1994a,th. Thus, KanCOs play a cardiopresective role, whereas glibenclamide worsens myocardial stunning (Auchampach et al., 1992). Analogies have been observed between the cardiopentection conferred by KanCOs and ischemic preconditioning (i.e., increased tolerance of cardiac myocytes to an ordinarily lethal ischemic insult, achieved by an initial brief exposure to ischemia), for example, both are sensitive to glibenclamide blockade (Gross and Auchampach, 1992). Thus, therapy with KATPCOs may afford a permanent 'chemical preconditioning' that confers on the heart the ability to better withstand transient oxygen deprivation and, consequently, to suffer less tissue damage during acute myocardial infanction:

Whether KanCOs exect their beneficial effects on the ischemic heart by a direct (myocardial) or indirect (vascular) action remains to be determined. Studies with U-89,232 (cromakaline analog; Toxonbs et al., 1992) and BMS 180448 (pinacidil analog; Grover et al., 1995), KatriCOs devoid of vascular effects demonstrated cardioprotection against ischemia in animal models preater than that observed with cromakalim. This would suggest that there is a direct myocardial action of these second generation KariCOs that will provide an opportunity to explore the cardioprotection of such agents without possible complication (e.g., hypotension, coronary small. The mechanism of tissue selectivity is not clear, but it may be related to the existence of KAICO 'receptoe' subrypes in different tissues (Atwal, 1994). Similar concepts of subtypes of the site(s) of action of KarrCOs in SMC previously have been proposed in SMC (Piper et al., 1990; Wickenden et al., 1991; Lawson and Hicks, 1993).

Although Kary channel opening in response to ischemia may offer a cardioprotective mechanism, there is another consequence. The resulting increase in K' efflux shortens the action potential duration and contributes to the extracellular K* accumulation observed during an ischemic episode (Coerzee, 1992). These changes in conductative and extracellular K+ have been hypothesized to be responsible for ischemia induced arrhythmias. KariCOs have demonstrated arrhythmogenic properties in certain animal studies (Chi et al., 1990; Tosaki et al., 1992; De La Coussaye et al., 1993). However, although KAHCOs may be contraindicated in some types of arrhythmia, they may be of use in the treatment of certain other types of arrhythmia resulting from a repolarization defect (Antaelevirch and D. Diego, 1992). KauCOs have been shown to suppress thythm abnormalities related to delayed repolarization and early afterdepolarizations in anaestherised rabbits (Carlsson et al., 1991).

4.1.3. Blood. The K_{AIP}COs, cromakelim, celikalim, and pinacidil, inhibited white rhrombus formation in a rabbit arteriovenous shunt model, although they had no effect on human plateler aggregation (Parelunas-Hoffman et al., 1994). Antichrombotic activity of K_{AIP}COs in two may be related to hereficial effects on blood rheology and reduced red blood cell deformability.

4.2. Respiratory System

Administration of cromakalim and other KarsCOs to conscious (oral or inhalation route) or anaesthetized (oral, inhalation or i.v. route) guines-pigs protects against histamine, 5-HT, or (in sensitized animals) availbumin-induced bronchoconstriction (Raeburn and Karlsson, 1991). In the anaesthetised animal (Konzert-Rossler model, where protective cardiovascular reflexes are inhibited), a reduction in

diastolic blood pressure was observed following oral or i.v. administration of KAISCOs. Bronchodilation, however, could be achieved at doses of cromakalim not reducing mean arterial blood pressure in experiments where the KATPCO was administered by inhalationly thus, demonstrating selectivity (Bowring et al., 1991; Raeburn and Karlsson, 1991; Bowring et al., 1993; Arch et al., 1994). Respiratory dynamics measurements in anaesthetised guinea-pigs revealed that the KARCOs, cromakalist and BRI, 55834, resembled theophylline by eliciting similar inhibition of histamine induced increase in airways resistance and decrease in lung compliance, and salbutamol, a B-agonist, was more effective against resistance than compliance (Bowring et al., 1991). Therefore, KarpCOs, compared to Bagonists are more effective dilators of small airways (where constriction decreases compliance) for identical large airways effects (where constriction increases resistance). Reports have implicated the activation of K_{Cs} channels in the relaxant responses of respiratory smooth muscle to Biagonists (Kume et al., 1989; Jones et al., 1990). Thus, these findings may be suggestive of the distribution of KATP and KGs channels within the smooth muscle of the respiratory system. The clinical relevance of such a hypothesis is, as yet, unclear,

KANCO can inhibit neurotransmitter release from cholinergic and nonadrenergic, noncholinergic excitatory neurones (NANCe) in guinea-pig lung in vitro and in vivo (Raeborn and Karisson, 1991; Small et al., 1992). These neural effects of KarrCOs may be very relevant to potential reasment of asthma because, not only does bronchoconstriction frequently have a significant parasympathetic component, but also neurogenic inflammation of the long may contribuse to the pathology of asthma (Batnes et al., 1991). The main evidence for an effect on neurotransmitter release is that KapCOs are far more effective at inhibiting cholinergically or NAMCs-mediated broncheconstriction or mucus secretion elicited by stimulating neurotransmitter release than when the relevant neurotransmitter is supplied directly (Ichinose and Barnes, 1990; Burka et al., 1991; Raeburn and Karlsson, 1991; Small et al., 1992). A prejunctional site of action has been proposed for the inhibition of peptidergic excitatory neurotransmitter release due to KADCOs inbibiting the NANCe neuroeffector transmission at concentrations slightly lower than those causing relaxation of airways smooth muscle. Interestingly, the KanCOs do not seem to interfere with NANC inhibitory neuroeffector transmission in the lung (Burka et al., 1991). Nielson-Kudsk et al. (1994) demonstrated that, like cromakalim and pinacidil, terbutaline-(B-agonise), theophylline-(xanthine), and verapamil-(Cal+ antagonist) induced inhibition of NANCe neuronmusmission in guinea-pig bronchi involved a prejunctional sire.

The inhibition by cromakalim of electrically evoked [41]acetylcholine release in rat isolated trachea has been suggested to be an epithellum-dependent mechanism (Wessler et al., 1993). This effect was only observed in tube preparations, where the mucosal/submucosal environment would be better preserved, and not in trachea opened longitudinally. KAPPCOs. BRL 18227 and YM-934, inhibited plasma leakage in traches, main bronchi, and central and peripheral intrapolmonary airways evoked by stimulation of vagal nerves in guinea pigs (Lei et al., 1993; Ishikawa et al., 1994). These compounds had no effect on exogenously administered Substance P-induced plasma leakage. Thus, cromakalim and YM-934 inhibit airway neurogenic inflammation by modulating the release of neuropeptides from the sensory nerve endings, and the inhibitory effect can be attributed to the KCO activity.

KATECOs can reduce obstruction to airflow by suppressing hyperreactivity of intact airways in animals, with doses that are insufficient to relax strway smooth muscle in situ in normal animals (Chapman et al., 1991; Paciorek et al., 1992; Morley, 1994). Hence, the potency of KarpCOs as inhibitors of bronchospasm is greater in hyperreactive than normal animals. An almost universal characteristic of asthmatics is that their airways are hyperresponsive to a wide range of physiological and pharmacological stimuli (Smith, 1992). The causes of airways hyperresponsiveness in humans are not well-defined, although several animal models have been developed to emulate this response. In general, however, the degree of hyperresponsiveness achieved in animals is less than that in humans (Smith, 1989). Compounds that open K1 channels and impair expression of airway hyperreactivity in the absence of direct smooth muscle spasmolysis will provide a novel approach to symptomatic therapy in aschma (Mocley, 1994).

In general, the direct relaxant properties of KarrCOs such as cromakalim have been assessed predominantly in guinea-pig isolated trachealis moscle (Rachuen and Karlsson, 1991, Longman and Hamilton, 1992, Small et al., 1992). This vissue has also been used for ion flux studies and for intracellular recording of change in membrane potential. KateCOs inhibit contractions to or reverse precontractions to a variety of spasmogens in guinea-nig tracheal prepatations. However, in interaction studies, the smooth muscle relaxant responses to KAnCOs in the guinea-pig trachea are blocked by glibericlamide in a manner that is not consistery with competitive antagonism (i.e., the maximum effect of the KarrCO is reduced in the presence of the antagonist (Berry et al., 1991; Nielsen-Kudsk et al., 1990)), but is consistent with a lack of spare channels (i.e., the $\mathsf{K}_\mathsf{AU}\mathsf{CO}$ must activate all channels in the tissue to evoke a maximal response; see Section 2.1). The arragonism by glibericlamide of the effects of KenCOs in functional in vitro pharmacology studies on vascular smooth muscle and certain nonvescular smooth muscle preparations has been described as competitive in nature (see Section 2.1).

The lack of competitive interaction between glibenclamide and the KAPPCOs is indicative of the involvement of more than one mechanism in the relaxant effect of the latter. This is consistent with the conclusion that relaxations of tracheal smooth muscle to BRL 55834 (a benzopyran KAPCO) is mediated by, at least, a glibenclamide-resistant K* channel (Lawson et al., 1993s; Edwards et al., 1995). In addition, BRL 55834 has

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been reported to activate an ATP- and glibericlamide-sensitive K* channel and, at higher condentrations, a large conductance charybdotoxin-sensitive Ca**-activated K* channel in bovine trachealis SMC (Ward et al., 1992).

Cromskalim and levrocromakalim have demonstrated relaxant effects in human bronchial smooth muscle (Taylor et al., 1992; Black et al., 1990). Differences (primarily in potency) from the findings obtained in gumes-pig preparations suggested that the guines-pig is not a good predictor of the inhibitory response of KATCOs in humans.

Clinical trial of cromakalim in patients with nocturnal assibma showed that a single (0.3 mg) or repeat (0.25, 0.5 mg) oral dose administered at 11:00 p.m. could attenuate the dip in lung function measured at 6:00 a.m. the following morning (Williams et al., 1990). The predicted peak plasma concentration of cromakalim in these studies was about 5-fold less than its threshold concentration for relaxation of tone in human bronchi (Yaylor et al., 1992). Studies in animal models prior and subsequent to these findings suggest that the positive results are due to actions other than just relaxation of the broachial smooth muscle (Longman and Hamilton, 1992; Small et al., 1992; Morley, 1994). The efficacy of cromakalim may not involve the direct relaxation of airways smooth muscle, but is due to an influence on neural mechunisms underlying airways hyperresponsiveness. This suggestion is supported by the effects of KABCOs in animal models of hyperreactivity (Morley, 1994); the potency of cromakalim in human hyperresponsive anways smooth muside is, as yet, unknown.

Interestingly, when the dose of cromakalim was increased to 1.5 mg (single dose), no significant reduction in the morning dip in lung function was observed (Williams et al., 1990). The failure of the latter dose of cromakalim to improve long function was attributed to 10 out of the 23 subjects being unable to exert maximal expiratory effort during measurements of FEV3 values (forced expiratory volume in 1 sec) as a consequence of headache, BRL 38227, which replaced cromakalim in clinical relats, failed to elicit significant bronchodilation or reduce bronchial hyperresponsiveness to histomine or methacholine when administered as a single oral dose to asilymanics (Kidney et al., 1993); rhus, not meeting the criteria set for its development in asthma. As with cromakalim, the dose-limiting side effect of oral BRL 38727 is headache, probably resulting from cerebral vasodilation (Arch et al., 1992). Bimakalim, an analog of cromakalim, also lacked bronchodilatory effects following inhaled adminserration to mild to moderate broachial asthmatic adult paraches (Fadschees et al., 1994). Whether this was a rive fack of broachial efficacy or that the dose of drug, to avoid other effects (no headaches or cardiovascular effects reported), was too low requires further investigation. Therefore, to be useful ocal bronchodilators, and have the potential to reduce bronchial hyperresponsiveness, KarrCOs with selectivity for airway relative to vascular smooth muscle greater than that of cromakalim or BRL 38227 are required.

BKCs have been demonstrated in high density in canine, boving, and human virway smooth muscle (McCanti and

Welsh, 1986; Green et al., 1991; Miura et al., (991). This has led to the proposal that openers of BKCs channels could demonstrate therapeutic benefit in regulation of one of the respiratory system. The findings involving BKCs channels in the effects of \$\mathcal{B}\$-agonists on airways smooth muscle (Kume et al., 1989; Jones et al., 1990) support this hypothesis. In human bronchi, however, NS 1619 (a BKCs channel opener; Olesen et al., 1994a,b) evoked a weak relaxant effect that did not involve BKCs channels, which could call into question the utility of such drugs as bronchoddators (Templeton et al., 1995). The lack of selectivity of KCsCOs for the BKCs channel (see Section 2.2) has delayed progress in this area and, thus, appreciation of the true utility of such agents.

4.3. Reproductive System

By virtue of the smooth muscle relaxing effects, Kap channel openers may be useful in the treatment of premature labour and dysmenorthoes (Piper et al., 1990). Several KarrCOs are capable of producing glibenciamide-sensitive relaxation of uterine smooth muscle of car, both in vitto and in view (Piper et al., 1992). Cromakalim inhibited the spontaneous phasic activity and spasmogen-induced contractions of isolated uterus from the term-pregnant rat (Hollingsworth et al., 1987). BRL 38227 and phracidil labibited spontaneous and exytochr-induced contractions in human isolated pregnant myometrium, obtained before and after the onset of Intour (Cheuk et al., 1993); Morrison et al., 1993). The KAHCOs were more potent in nonpregnant than pregnant human myometrium (Cheuk et al., 1993). The relaxant effects of the two KareCOs in human pregnant myometrium was sensitive to glibenclamide. Thus, K_{ATP}COs may have potential as a new generation of toxolytic agents. Preferential higher potency would suggest that KareCOs would be more effective tocolytic agents in nonpregnant than pregnant women (Check et al., 1993). Not all women with preserm oterine contraction, however, are candidates for recolvais (Monga and Creasy, 1995),

Although channels permeable to Rh* and K* exist in the uterus, cromakalim does not stimulate efflox of these ions in rat uterus, even though relaxant responses were sensitive to glibericlamide (Hollingsworth et al., 1987). This finding was further supported in human isolated myometrium, where Rh* exhibited differentiation on the effects of BRL 38227 and P1060 (a pinacidil analog) on amplitude and frequency of sponraneous contraction (Criddle and Soares de Moura, 1995). In these studies, Rh*-sensitive and Rh*-insensitive mechanisms were identified, of which the former appeared more important in effects of P1060 than BRL 38227. Although KANCOs may demonstrate benefit in uterine-associated disorders, the involvement of K* channels still requires confirmation.

B-Adrenoceptin agonists, relaxin, and other uterine relaxants that increase intracellular cAMP levels, activate K_{Ca} channels in myometrial cells (Trithart et al., 1991; Meera et al., 1994; Sanborn, 1999). Iberiotoxin, a K_{Ca} channel blocker, depolarizes myometrial cells and increases phasic

contractions in rat and human myometrial preparations (Anwer et al., 1993). Therefore, selective direct openers of K_{Cs} channel (iberiotoxiti-sensitive) may also have therepeutic benefit in uterine-associated disorders.

Interestingly, DHS-I, derived from the medicinal bath Desmodium adscendent and used therapeutically in the treatment of dysmenorthea (Ampofo, 1977), opens charybdotoxin-sensitive K_{Ca} channels (McManus et al., 1993).

During labout, the forceful contractions of the uterus can occlude its blockl supply, which may lead to hypoxia. Hypoxia can reduce and even abolish uterine force in both isolated tat and human uterus (Heaton et al., 1993). Thus, hypoxia may contribute to uterine dystocia (inadequate uterine contraction during labour), the cause of which remains largely unknown and which often results in emergency Caesarean delivery. In a model of hypoxia involving cyanide administration to inhibit uterine force, glibenclamide reduces the K* efflux produced by cyanide (Heaton et al., 1993), thus implicating KATE channels in this response.

More work is needed to increase our understanding of the K⁺ channels in the myometrium; how they may differ from those in other rissue types and how they may be involved in different pathologic states.

in the treatment of impotence, vasodilators, injected locally, are commonly used. The established vascular smooth muscle relaxant properties of KADCOs (see Section 4.1.2) suggest that these drugs may offer an alternative to current creatment. Cromakalim and pinacidal increased *Rb+ efflox and inhibited spontaneous contractile activity, electrically and noradrenaline-induced contractions in rabbit isolated cavernosal rissue (Holmquist et al., 1990a). Similar results were obtained for pinacidil in human isolated cavernosum (Holmquist et al., 1990b). In addition, pinacidil increased whole cell X1 current in human cultured corporal (corpus cavernosum) SMC (Christ et al., 1993). Recently, cromakalim was reported to increase intracavernous pressure in a Simian mankey model, resulting in an erectile response of the penis (Trigo-Rocha et al., 1994). Minoxidil was more effective in facilitating election and produced fewer side effects than microglycerin, when used to treat organic imporence in men (Cavallini, 1991).

4.4. Urinary Bladder

Bladder hyperactivity, secondary to bladder hypertrophy or partial outflow obstruction resulting in urinary incontinence, is common, and the existing therapeutic regimens are often ineffective or poorly relevated (Wein, 1991). Cromakalim and pinacidil relax urinary bladder smooth muscle, indicating potential in the treatment of urinary incontinence (Andersson et al., 1988; Malmgren et al., 1989). In isolated detrusor muscle preparations from human unstable bladder (due to urinary outflow observation), cromakalim inhibited elevated basal tone and spontaneous contractile activity (Poster et al., 1989). Similar effects were also observed with pinacidil (Povaeus et al., 1989). KatpCOs demonstrated inhibitory effects, in both human and animal rissues, on myogenic activ-

try and contractions to a variety of spasmogens (Longman and Hamilton, 1992). The ability of KAIPCOs to inhibit electrically induced contractions of urinary bladder tissues have been variable and may be related to the degree of depolarization produced by neuronal stimulation in different models.

In bladder rissues from a variety of species, K_{AD}COs (e.g., BRL 38227, pinacidil and RP 49356) increase ⁴²⁴⁴K* efflux. However, as found in other smooth muscle systems, the concentrations of drug required are higher than those inhibiting myogenic activity (Longman and Hamilton, 1992). The relaxant activity and enhancement of ⁴²⁴³K* efflux due to K_{AT}COs are sensitive to glibenclamide.

Evaluation of KarrCOs on in visc urinary bladder models have been limited due to the viscular effects of the drugs. A comparison of the effects of BRL 36227, pinacidil, Ro 31-6930, RP-49356, and S 0121 did not reveal selectivity for the randerrusor muscle over portal vein (Edwards et al., 1991). A series of novel KarrCOs (for example, compound ZD6169) recently have been proposed that act selectively on the urinary bladder smooth muscle, without producing significant cardiovascular effects following oral administration to rats (Grant et al., 1994).

The observations in animal models have yet to be supported by clinical trials, as results of initial studies in humans were disappointing. Data from a study with pinacidil in patients with bladder hyperactivity and bladder ourflow obstruction (secondary to prostatic hyperplasia) failed to demonstrate any clear improvement in bladder function (Hedlund et al., 1991). Levrocromakalim (BRI, 38127) increased the duration of bladder contraction, but was without effect on other utodynamic parameters in patients with high spinal cord lesions (Komersova et al., 1995). Hypotensive responses during this study led the authors to suggest that higher doses of the drug could only be evaluated if administered intravesically.

The relaxant effects of cromakalim and \$ 0121 (bentopytan K_{ATT}CO) in isolated uteter from rabbit and humans (Klaus et al., 1990) suggest a benefit for K_{ATC}COs in the reatment of kidney stones by alding their passage along the uterer (Englert et al., 1988). In studies on the guinea-pig uteter, CORP (see Section 2.3) appears to be an endogenous KCO (Santicioli and Maggi, 1994). The role of CORP and the involvement of K^{*} channels on human uteter still requires investigation.

4.5. Gastrointestinal Tract

Spontaneous slow-wave contractile activity and/or contractile responses to spasmogens in a variety of gastrointestinal tissues (e.g., taenia caeci, tleum, colon, muscle myenteric plexus, oesophageal, stomach) have been shown to be inhibited by KathCOs (Longman and Hamilton, 1992). These effects were associated with K* efflux and hyperpolarization that involved a glibenclamide-sensitive mechanism. The animal data suggest that KathCOs may have utility in conditions associated with disturbances in gastrointestinal motil-

ity, such as itritable bowel syndrome, especially because kinetically slow K* channels, carrying ourward current, may be responsible for gastrointestinal slow-wave activity (Benham and Bolton, 1983). Interestingly, the evaluation of such drugs in other clinical conditions has not revealed an incidence of adverse side effects on the gastrointestinal tract such as constination; however, this would be influenced by the site of adsorption of these agents. To gain a full appreciation of potential therapeutic benefit, KCOs are required that are not removed from the gastrointestinal tract.

Diazoxide, like morphine, has a protective effect on ethanol-induced gastric lesions, an effect proposed by Bhounsule et al. (1992) to involve K_{ATV} channels. As described in Section 4.6.2, opioid receptors are associated with K* channels, and glibenclamide is reported to antagonise morphine analysisis (Ocana et al., 1990). In the model of gastric lesions, results obtained with glibenclamide were complicated by its action on prostaglandin production in this tissue (Bhounsule et al., 1992).

Neurorensin inhibits contractions in rat and carrine ileal smooth muscle by opening apartin-sensitive K_{Cs} channels (Allescher et al., 1992; Christinck et al., 1992). Therefore, SK_{Cs}COs, like K_{AD}COs, may have utility in conditions associated with disturbances in gastroinrestinal motility.

4.6. Nervous System

4.6.1. Peripheral. The KATICOs interfere with neuro-transmission in peripheral parasympathetic neurones in the airways and the gastrointestinal tract (Longman and Hamilton, 1992; Section 4.2). This has led to suggestions of a presymaptic site of action, whereby the KATICOs control the release of neurotransmittet. In contrast, cromakalim, nicorandil, and pinacidal failed to exert an inhibitory effect on nonadrenaline release in rat isolated mesenteric artery (Fabiani and Story, 1994). The presynaptic inhibitory role of KATICOs, therefore, appears to be selective for parasympathetic (cholinergic) innervation.

Interestingly, cromakalim and pinacidil inhibited nicotinic acetylcholine receptor mediated and voltage-dependent catecholamine secretion from bovine adrenal chromaffin cells (Masuda et al., 1994). Thus, KanCO-sensitive K* channels could be involved in regulation of catecholamine secretion mainly indirectly through VOCs.

4.6.2. Central. Potassium channels play a pivotal role in the control of neuronal excitability, action potential, and neurotransmitter release within the CNS (Hille, 1984; Cook, 1990). Activation of a variety of receptors (e.g., opioid, 5-HT), somatostatin, αγαdrenoceptors) by the appropriate neurotransmitter alters the flux of K* ions from neurones (North, 1989). Because of this role in normal CNS physiology, decongements in the function of K* channels may underlie several CNS diseases. Studies of the distribution of binding sites for the three ligands, [12]-iodoglyburide, [12]-apamin and [12]-charybdosoxin, as markers for Kare, SKCs, and BKCs (or voltage-gated K*) channels, respectively, was deamstically different in the rat brain (Gehlert and

Gackenheimer, 1993). These data indicate that pharmacological modulation of these K* channel subtypes should result in distinctly different effects on brain function. [25]}lodoglyburide binding exhibits a very broad distribution in basin, being found in a majority of brain regions. The globus pallidus and the zona reticulara of the substantia nigra linvolved in movement coordination) contain the highest density of binding. Openers of KATE channels have been suggested to exert a protective effect on ischemic tissue by reversing the ischemia-induced depolarization (see below). Therefore, KarrCOs may find utility as neuroprotectant drugs, and the broad distribution of these channels would indicate that these drugs would have an effect on most neuronal populations in the brain. Although KareCO ligands, PHP1075 and PHloromakalim, are available, a profile of the binding site(s) distribution within the CNS is not published as yet. [331] Apamin binding suggested that SKG channels are associated primarily with cell bodies and dendritic spines, rather than nonneuronal elements. Localization in the cerebral cortex and hippexampus would suggest that this channel may have a role in the processing of memory. Interestingly, a loss of [23] apamin binding sites has been reported in the subjection and CA; neurones of hippocampal tissue obtained from posimortem Altheimer's disease patients (Ikeda et al., 1991). A reasonable hypothesis is that openers of the 5KC, channel may confer a neutoprotective effect, however, development of appropriate chemical molecules and pharmacological tools is awaited. The highest levels of [123]-charybdoxoxin binding sites were found in the white matter-containing regions, such as the lateral offactory tract and fasciculus retroflexus. This suggests that the charybdotoxne-sensitive K* channel is present on agons and may modulate herve conductance in these regions.

Studies in vitre and in animal models indicate the potential clinical utility of KarpCOs for diseases of the CNS.

During anoxic conditions, neuronal depolarization is due, at least, to the release of large concentrations of excitatory amino acids, such as glutamate, which may be involved in long-term is chemic-induced damage to the brain (Miller, 1990). In in view experiments, dissocide and somatostatin were shown to prevent anoxia-induced depolarization of CAB hippocampal agusons following the opening of K+ channels (Ben-Ari et al., 1990); these effects were inhibited by precreatment with glyburide. The authors proposed that KarpCOs may prevent anoxia induced damage to hippocampal neurons by inhibiting the release of excitatory amino acids. This suggestion is supported by the finding that levrocromakalim and RP 52891 blocked ischemia-induced glutamare release in for hippocampal slices (Zini et al., 1993), in addition, the KanCOs levrocromokalim, nicerandil, and pinacidil blocked ischemia-induced expression of the genes cales and columnand of the mRNAs for 70-kDa hear-shock protein and the form of the amyloid \$-protein precursor, including the Kunitz-type protease liabilities domain in or hippocampus (Heausteaus et al., 1993).

The movement disorders associated with Parkinson's dis-

ease are due to a selective loss of dopaminergic neurons in the substantia nigra. The highest density of Kame, as judged by autoradiographic studies with [181] lodoglyburide, in the brain is found in the substantia nigra (Ochlert and Guckenheimer, 1993). Sulphonylures binding studies, however, provide essentially only indirect evidence of Kares. Sulphonylureas or increasing extracellular glucose increase the release of [H]-CABA from the substantia nigra, effects that are inhibited by KatiCOs (Schmid-Antomarchi et al., 1990). GABAergic pathways to other CNS structures (e.g., raphe nuclei) are also modified by KATPCOs. Schmid-Antomarchi er al. (1990) noted that the order of potency of the Kan COs (leveromakalim > nicorandil > cromakalim > diazoxide > pinacidil) was found to be different from that in either the pancreatic B-cell or in smooth muscle, possibly indicating a difference between the target K* channel in this brain region and that in other tissues. The Kare channel in neuronal rissue is not the classical (Type 1) channel found in pancrearic B-cells or heart, but a large-conductance nonrectifying version (Type 2: Ashcroft and Ashcroft, 1990).

The genesis and propagation of nonphysiologic electrical impulses are the hallmark of epilepsy. Thus, the byperpolarization (and restraining) of excitable cells through the opening of K* channels could demonstrate cherapeutic benefit in this setting. The KADCOs etomakalim and RP 52891 (aprikalim) reduced seizures in genetically epileptic rars (Candolfo et al., 1989) and pentyleneterrazole-induced seixures in mice are blocked by intracerebrovenericular administration of cromakalim, but not pinacidil (Del Pozo et al., 1990), in a dilutazem-induced model of tonic-clonic seizures in the rat, cromakalim completely inhibited both EEG and behavioural seizures, and penrobarbitone only prevented behavioural activity (Popoli et al., 1991). Katt COs counterser anexic hyperexcitability, but not 4-aminopytidineinduced epileptiform activity in the rat hippocampal slice (Marcia et al., 1994), suggesting such drugs might be useful in the treatment of seizures occurring in the setting of status epilepticus or cerebrovasculat disease. High concere crations of cromakalim inhibit electrically induced epilepciform discharges in guinea-pig hippocampus (Altheimer and ten Bauggencare, 1988). Finally, the antiepileptic drug carbamazepine increases potassium currents in rat corrical neurons (Zona et al., 1990). However, whether or not this property is linked to the pharmacological actions of the drug requires further investigation. A similar property (K r channel opening), which may be of clinical importance, has been reported to be exhibited by excarbatepine (Mclean et al.,

Opioids exert their analgesic effects by binding to opiate receptors, which leads to opening of K+ channels and neuronal hyperpolarization (North, 1989). Morphine-induced antinociception in mice tail-flick tests is mediated by the opening of K_{ATP} channels (Ocana et al., 1993). These observations would suggest a potential role for KCOs as analgesics. Intrarhecal administration of the K_{ATP}COs BRL-38227, minoxidil, and distantide produced antinociception in the tail-flick test in mice (Welch and Dunlow, 1993). The

K_{AD}COs were not cross-tolerant to the effects of morphine in this model. This led to the suggestion that the K_{AD}COs and opioids probably do not act on a common site, but could have a common second messenger (Welch and Dunlow, 1993). A dose-dependent increase in the effects of morphine on the hot-plate and tail-flick rests were obtained following i.c., administration of pinacidil to rats (Vergoni et al., 1992).

Benzopyran derivatives of cromakalim (e.g., SR 46142A) have been claimed to exert antidepressant activity in animal tests (improve swimming performance in mice) in the absence of a cardiovascular effect (Garcia et al., 1990). Similar results were observed with a series of amidochromanols (Poucelet, 1990). This would suggest that modifications of the benzopyran nucleus of cromakalim allows tissue selectivity (CNS over vascular smooth muscle) to be achieved. Whether or nor the antidepressant effects of SR 46142A (and related compounds) are mediated by modulation of K* channels remains to be confirmed.

These studies are encouraging, but neuronal tissue-selective agents that cross the blood-brain barrier are needed to realize the potential clinical applications of KADCOs in CNS disorders. In addition, the promiscuity of KADCOs invites adverse CNS effects; for example, cromakalim, like d-amphetamine, enhances spontaneous locomoror activity in the rat by a glibenclamide-sensitive mechanism (Amalric et al., 1997).

BKCs channels also play a role in the function of newronal cells (Hille, 1984; Cook, 1990). The recent identification of compounds (e.g., NS 1619, see Section 2.2) that activare BKCs channels will allow determination of their utility as potential therapeuric agents. In rar cerebellar granule cells, NS-004 activates a 187 pS Ca-dependent K+ channel that is blocked by charybdotoxin (Olesco et al., 1994a). NS 1619 activates the BKCs, but not KATE channels, to membrane patches isolated from rar ventromedial hypothalamic neurones (Sellers and Ashford, 1994). Although Kary channels are present in rat ventromedial hypothalamic neurones, BRL 38227, cromakalim, and pinacidil all failed to evoke an effect on these channels in excised membrane parches (Sellers et al., 1992). The glucose-sensitive cells in the ventromedial hypothalamus are involved in the control of appetite and are often regarded as the satiety center (Morley, 1980; Blundell, 1992). Thus, acrivation of BKCs channels would decrease hyporhalamic firing and reduce the sensation of satisty.

4.7. Skeletal Muscle

In skeletal muscle, K_{ATE} channel activity has been shown to increase upon intracellular acidification (Davies, 1990). Falls in intracellular pH reduce the inhibitory effect of ATE on K* channels in frog skeletal muscle. This could mean that during increased muscle exercise and consequent lowering of pH, K_{ATE} channel-induced hyperpolarization could compensate for a decrease in electrical excitability and prevent spontaneous contractions from occurring.

The KarrCOs, cromakalim, pinacidil, and RP-49356,

increased opening time of a glibenclamide-sensitive K* channel in mouse skeletal muscle (Wesk and Neumcke, 1990). Discoxide activated Katt channels in smooth muscle and pancreatic cells, but had very little effect in skeletal muscle, even at very high concentrations (Weik and Neumcke, 1990). interestingly, this sulphonomide (thus, structurally related to the suphonylureas) inhibited ATP-sensitive channels in ventricular muscle cells (Faivre and Findlay, 1989). Thus, the different effects of diagoxide suggest that the K* channels in mouse skeletal muscle resemble those in cardiac cells more than those in smooth muscle and pancreatic B-cells (Lawson and Hicks, 1993). Due to diverse effects of pinacidil in mouse skeletal muscle, a model of the KATP channel was proposed with a binding site for the KanCO and rwo sites for nucleotides, one activatory and one inhibitory (ATP or ADP can occupy either site; Fiehl and Neumecke, 1994). Pinacidil activates the channel and displaces the blocker from the inhibitory site, only if the activatory site is occupied.

Cromakalim enhanced K* efflux in human skeletal fibres, an effect that was blocked by tolbutamide (Spuler et al., 1989), suggesting KATPCOs could have some role in the treatment of pathologic muscle fatigue or paralysis resulting from excessive membrane depolarization. Interestingly, cromakalim, pinacidil, and RP-49356 evoked larger hyperpolarizations in skeletal muscle fibres from patients with myotonic dystrophy or hypokalaemic periodic paralysis than in those from normal volunteers (Spuler et al., 1989; Quasthoff et al., 1989; Grafe et al., 1990). KATPCOs, however, increased open probability of an ATP-sensitive and an ATP-insensitive K* channel in human skeletal muscle (Quasthoff et al., 1990); the offect of the KATPCOs on both channels were blocked by glibenclamide.

Ischemia induced damage in a rat skeletal muscle, like cardiac (Section 4.1.1) and neuronal (Section 4.6.2) ischema, has been shown to be prevented by cromakalim (Hatton et al., 1991). This result, and the observation that cromakallim restores the membrane potential of depolarized human skeletal muscle fibres (Spuler et al., 1989), indicate that KatiCOs may be useful for the treatment of peripheral vascular disease (see Section 4.1.1). In rat skeletal muscle, Angersbach and Nicholson (1988) demonstrated that KATECOs, but not Call auragonists or hydralating, selectively increase blood flow to collateral vessels in a previously ischemic limb: Weselvouch et al. (1993), however, suggested that KATICOs would not be beneficial in treatment of skeleral muscle ischemia in vivo, but may be useful in preserving skeleral muscle function in cases of ischemia followed by reperfusion.

Studies in human muscle have implicated SK_{Ch} (apaminsensitive) charmels in the condition myotonic muscular dystrophy, characterised by muscle stiffness (Renaud et al., 1986). Alrhough K+ channel subtypes other than K_{AP} channels exist in skeletal muscle (SK_{Ch}, BK_{Ch}, delayed rectifier K+ charmel; Wareham, 1992), however, selective openers are awaited to determine their tole and the therapeutic potential of activation.

4.8. Hair Growth

The occurrence of hypertrichosis during antihyperrensive treatment with minoxidil (Campese, 1981) led to the subsequent evaluation of the drug (applied topically) to enhance hair regrowth in areas of baldness (Clissold and Heel, 1987). Topically administered minoxidit enhances hair growth in certain forms of male pattern baldness. Although this effect has been suggested to involve K* channels, hypertrichosis occurs in 80 -100% of minoxidit-treated patients, but only 2-13% of pinacidil-treated patients. There is no evidence that cromakalim, nicorandil, or RP-49356 stimulate hair growth. In SMC, minoxidil (sulphare) has been suggested to open a K+ channel that is not recognized by other KAUCOs (see Section 2.1). KAUCOS stimulate DNA synthesis in mouse epidermal kerarinocyte and whole hair folliele cultures (Harmon et al., 1993). In cultured whisker follicles, minoxidil, but not P1075 (pinacidil analog), preserved the root shearh; however, both drugs stimulated cysteing incorporation in follicles (Waldon et al., 1993). The root sheath may be the rarget for minexidil; thus, srimulating hair growth through a direct effect on the bair follide.

4.9. Intraocular l'ressure

Repeated topical application of piriacidil, cromakalim, or nicorandil lowers intraocular pressure (IOP) of rabbits, suggesting a potential benefit of K_{ATI}COs in eye disorders such as glaucoma (Godtfredsen, 1989). In an isolated arterially perfused bovine eye preparation, piriacidil caused a sustained decrease in IOP, with no effect on arterial perfusion pressure (Millar and Wilson, 1991); thus, suggesting that relaxation of resistance vessels is not involved in the fall in IOP due to the K_{ATI}CO. Whether these effects in the eye may be attributed to enhanced K* ion movement and consequent relaxation of smooth muscle or are the result of K* channel modulation in epithelial cells requires further investigation.

5. CONCLUSIONS

Synthetic molecules that 'directly' activate K* channels have led to a new direction in the pharmacology of ion thannels. The identification of the K+ channel opening properry of cromakalim initiated mojor research efforts in the search for other such agents and in the determination of the specific channells) involved. The existence of so many different subtypes of K* channels has been an imperus in the search for new molecules that would have different profiles and channel selectivities (e.g., Kgp, BKc,). The availability of an increasing number of KCOs, exogenous and endogenous, should facilitate more detailed study of these channels under both normal and certain pathophysiological conditions. The decrease in the excitability of cells that follows K . channel opening inters a broad clinical potential for drugs with this property in a number of pathologic conditions. Consequently, therapeutic roles of KCOs can be envisaged in disorders of a wide range of cells; for example, vascular and norwascular smooth muscle, cardiac, neuronal, and skeleral cells. Although lack of selectivity of current compounds remains one of the major hurdles in this area, advances in prototype KCOs and our knowledge of K* channel pharmacology is encouraging. Thus, the development of KCOs that will provide positive results in extensive clinical trials to give an appreciation of the full therapeutic potential are eagerly awaited.

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